

# MOD1 – DNA ENGINEERING

Fall 2010

**Day 2**

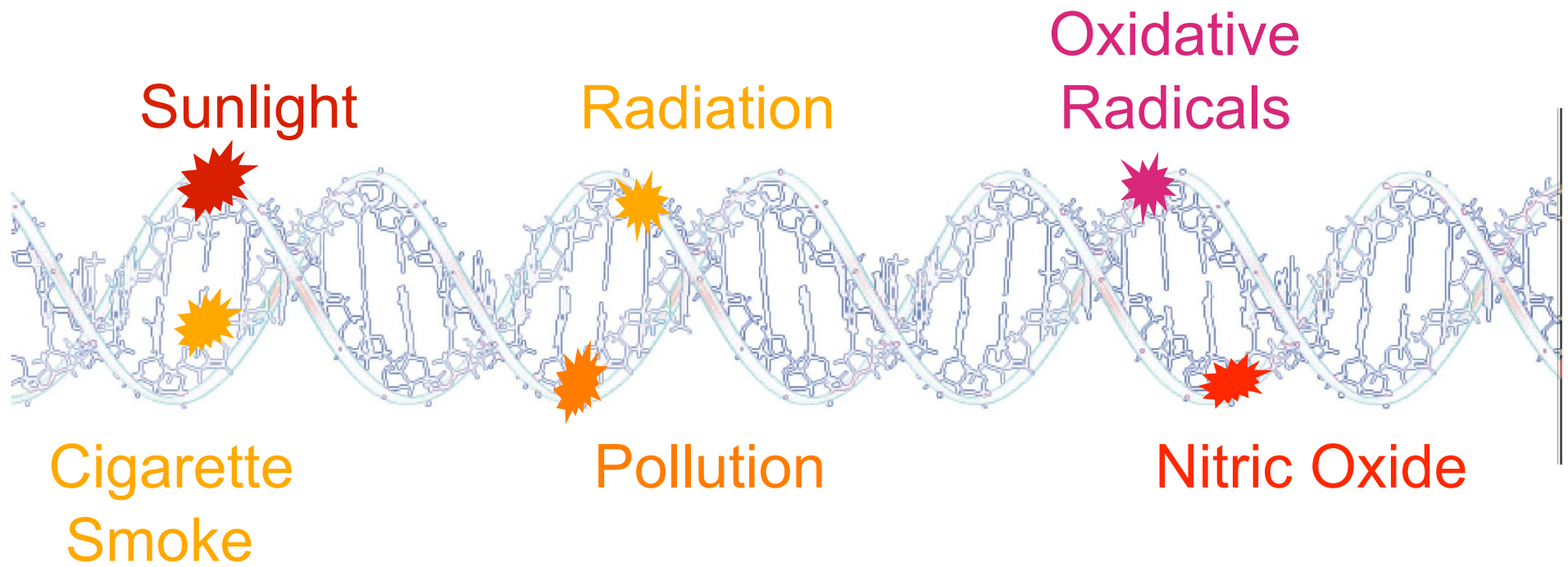
# **Mod1:**

**In this module, you will create a plasmid that will be used in an assay to measure homologous recombination activity in mammalian cells.**

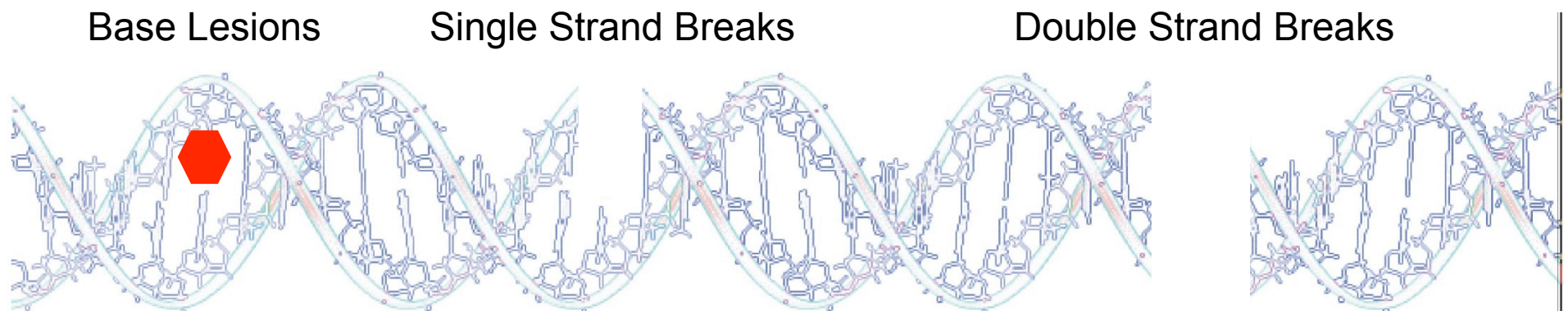
# Background & Significance:

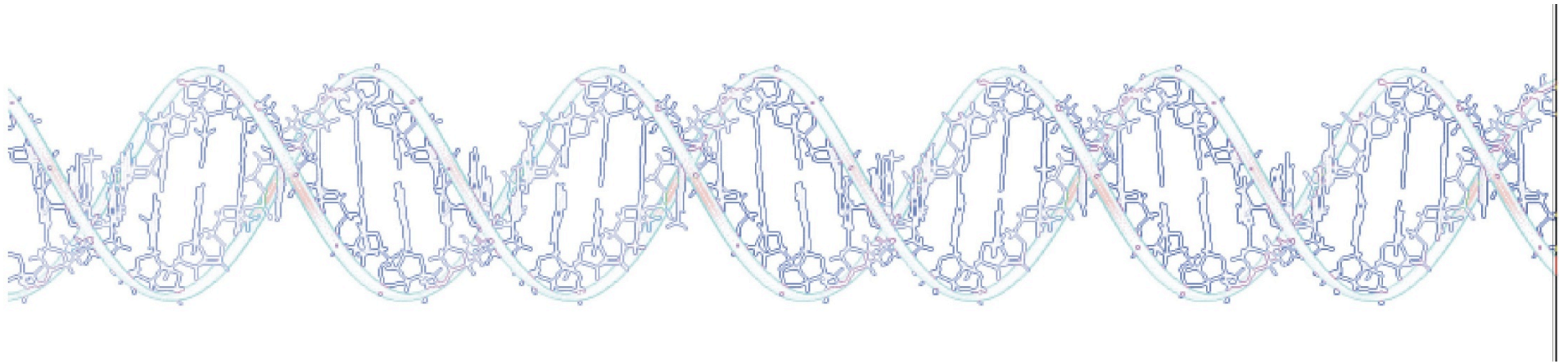
“Homology-Directed Repair” for  
double strand breaks

You will need to understand this material in  
order to analyze your data.



After damage, what might happen?





=



Double Stranded DNA



Sister Chromatid



Double Stranded DNA

Damage



Double Strand Break



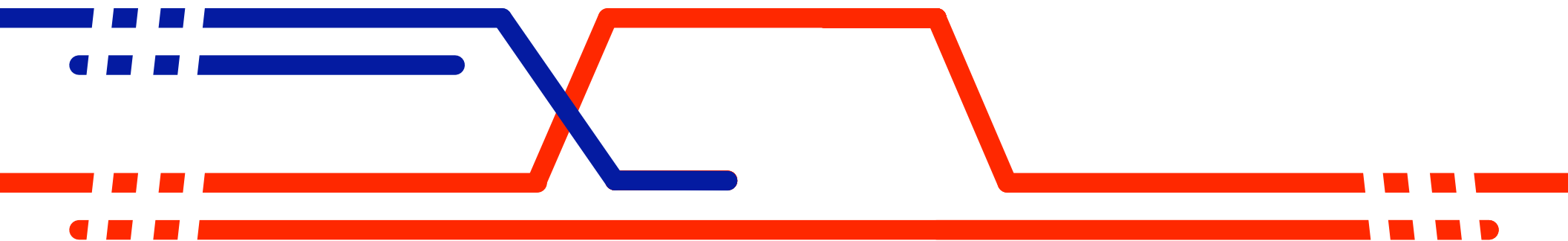
Resect to form 3' overhang



Find Homology

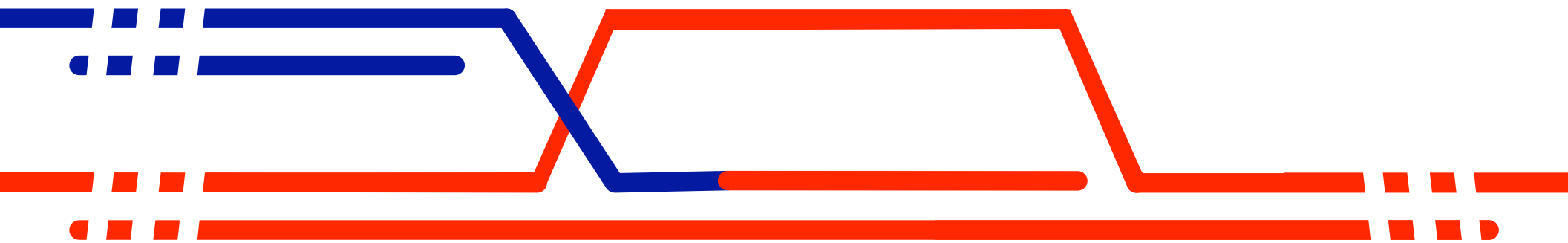


Form D-loop



Polymerize DNA using invading strand with 3'OH as a primer and the homologous donor DNA as a template





Branch Migration



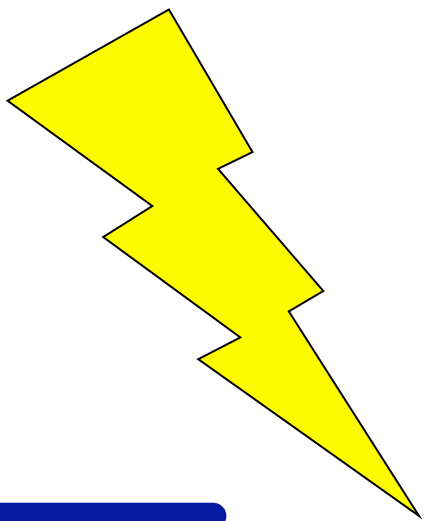
Bring extended ends together



Single Strand Annealing



Fill in gaps



+

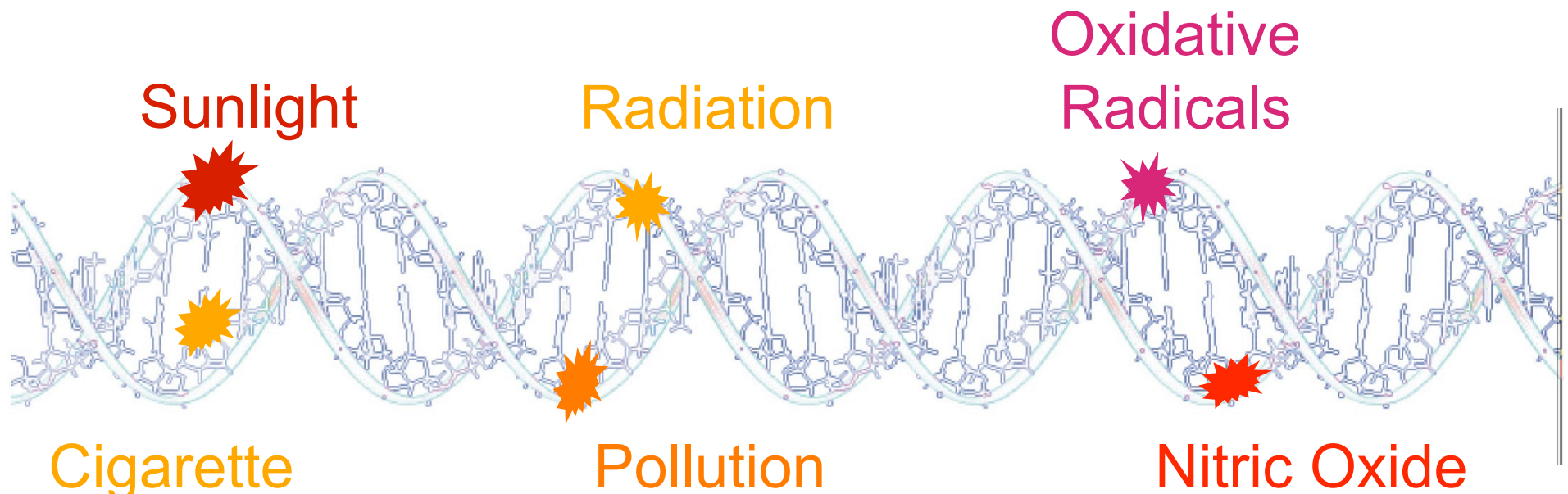


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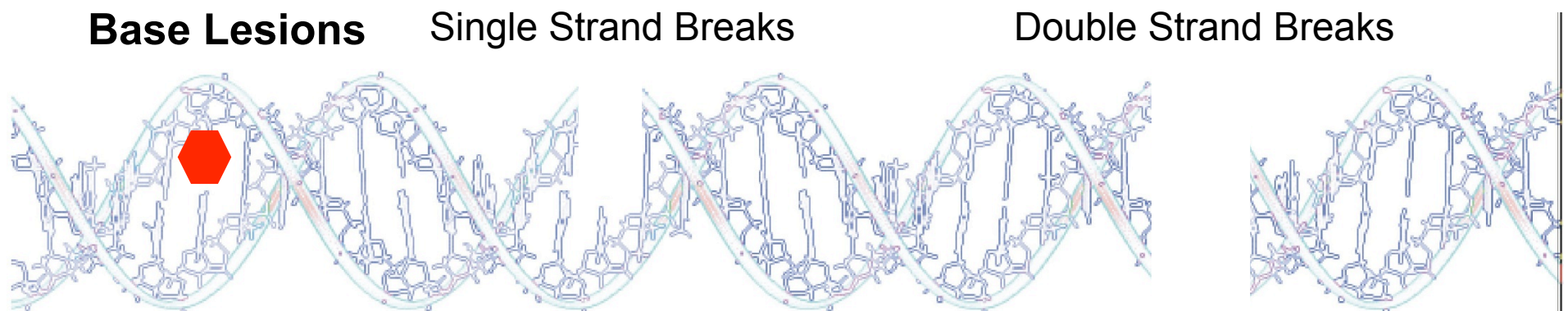


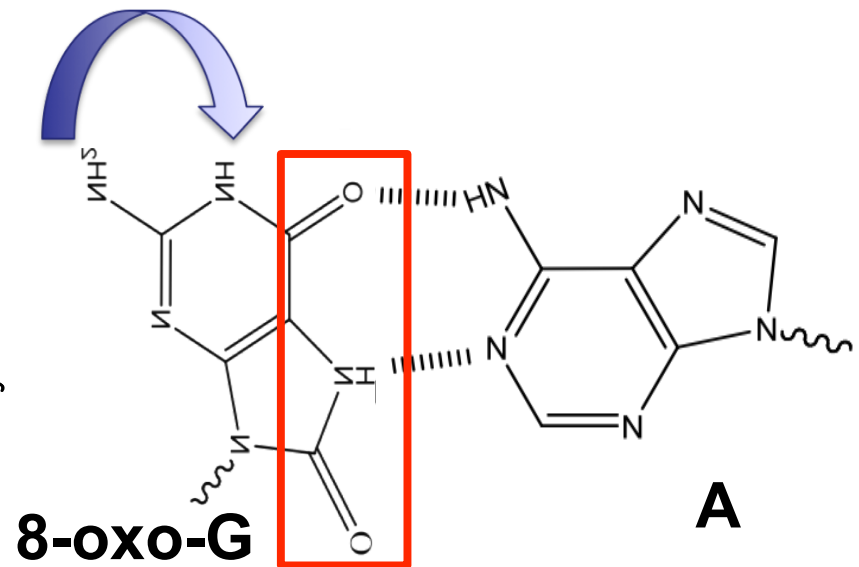
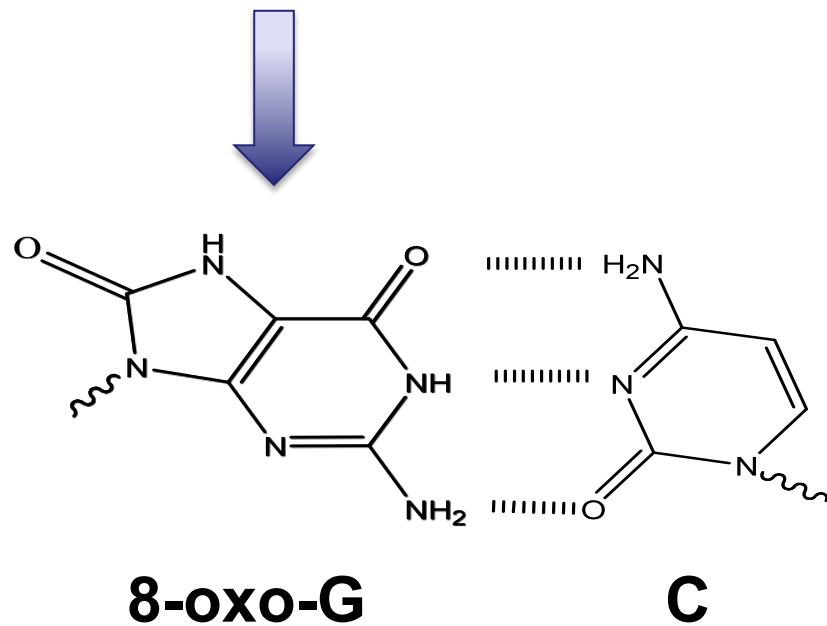
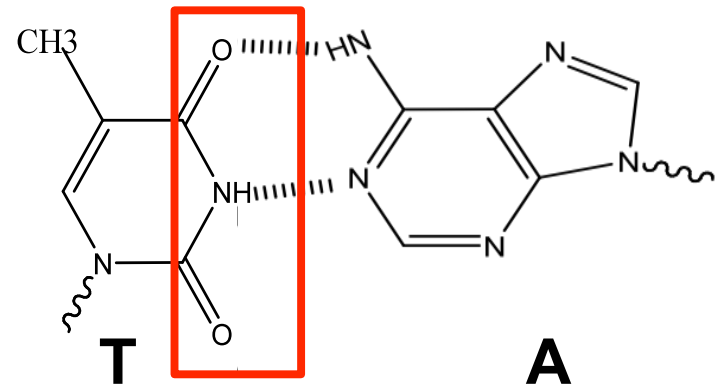
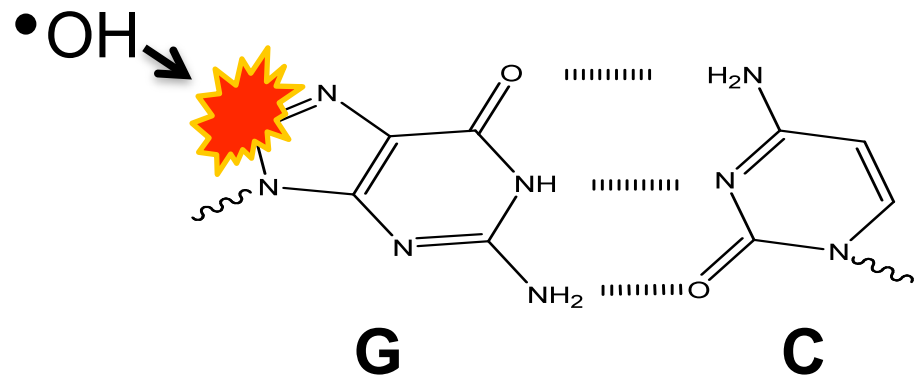
**See SDSA  
Prototypic Model  
& Replication Fork**

**Animations by Justin Lo**



After damage, what might happen?





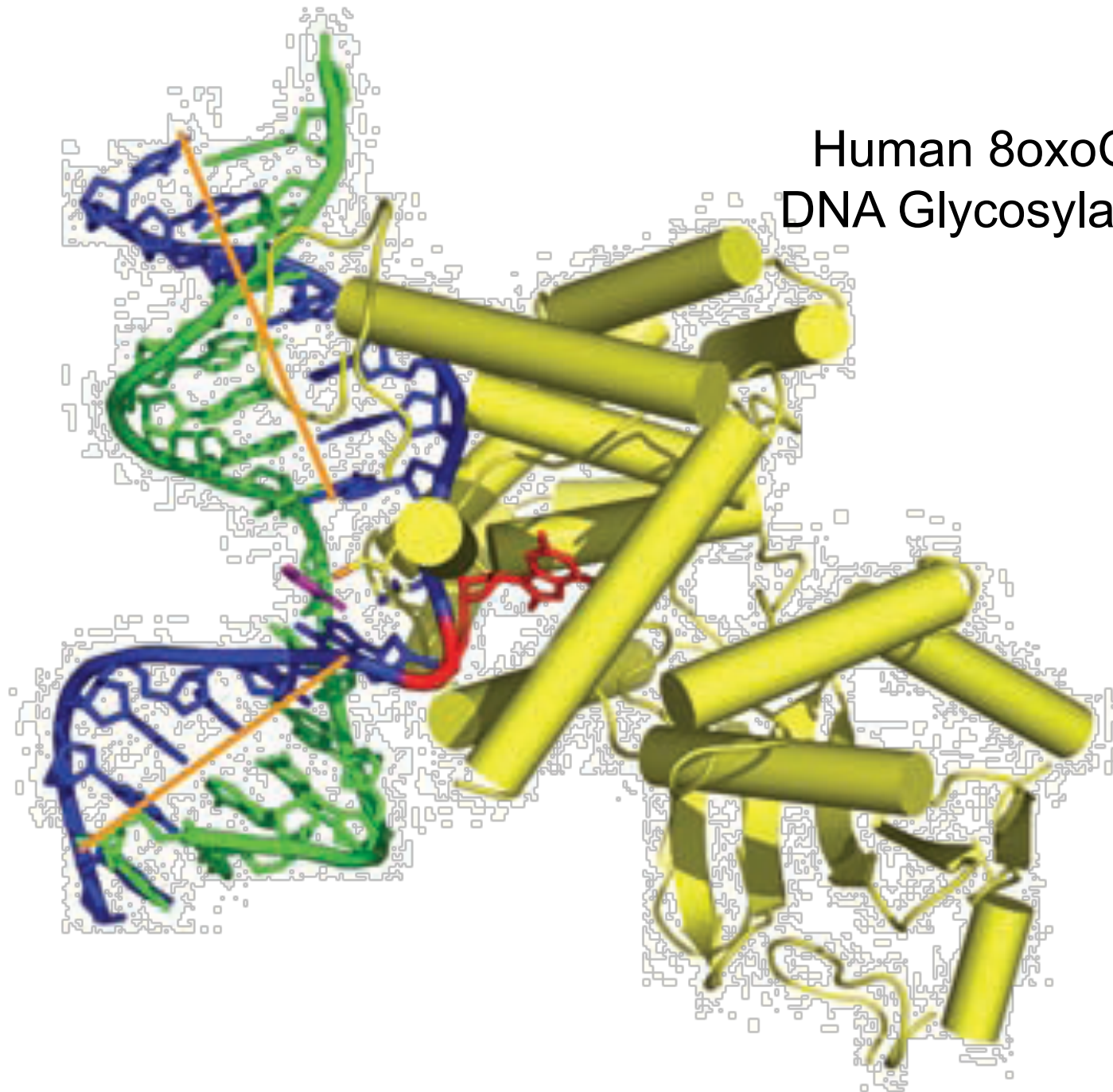
**GT -> AT**

# Does this damage matter?

- What can the cell do?  
Apoptose      Necrose      Nothing      **Fix it**
- If the cell is not dividing?
- If the cell is dividing?

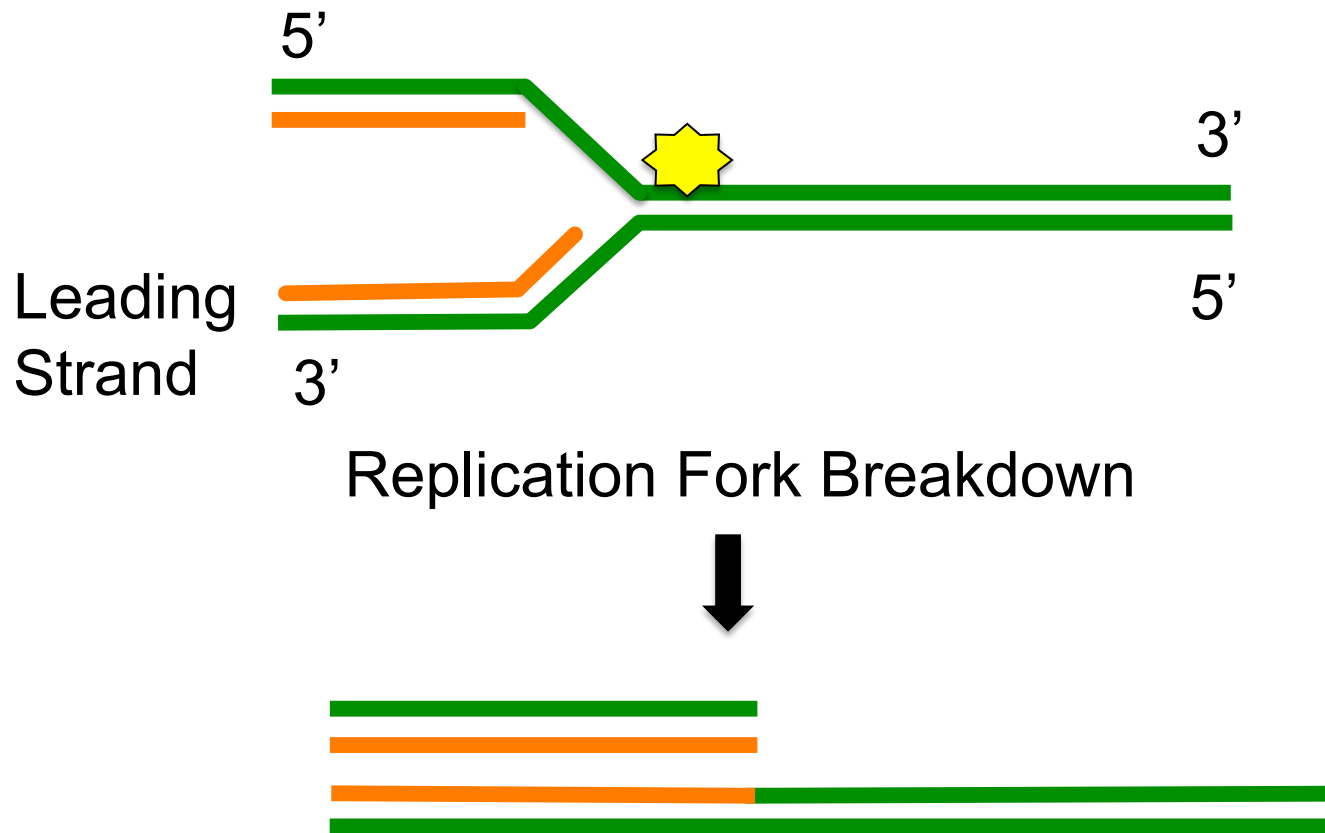


# Human 8oxoG DNA Glycosylase



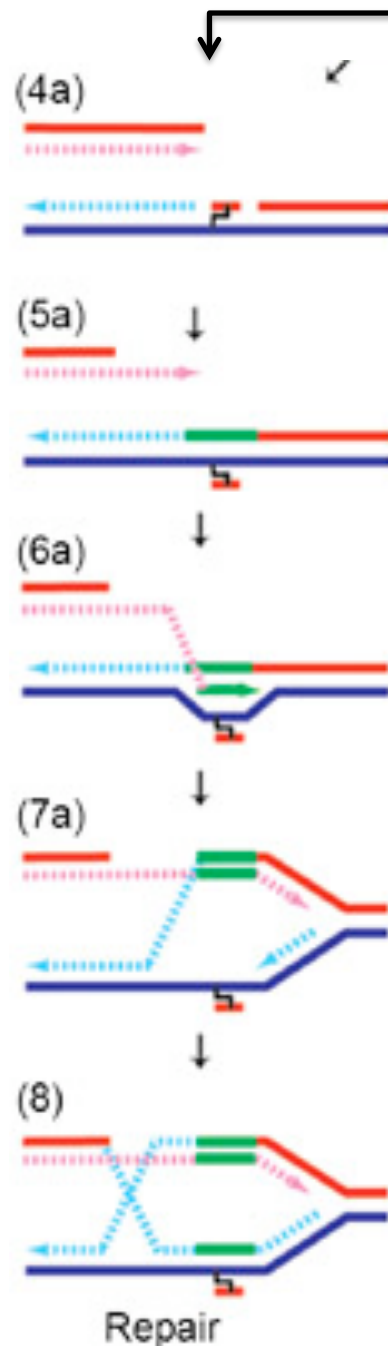
# Dangers of Bulky Damage

- Polymerase is unable to bypass lesions
- This leads to replication fork breakdown



# Homologous Recombination Videos

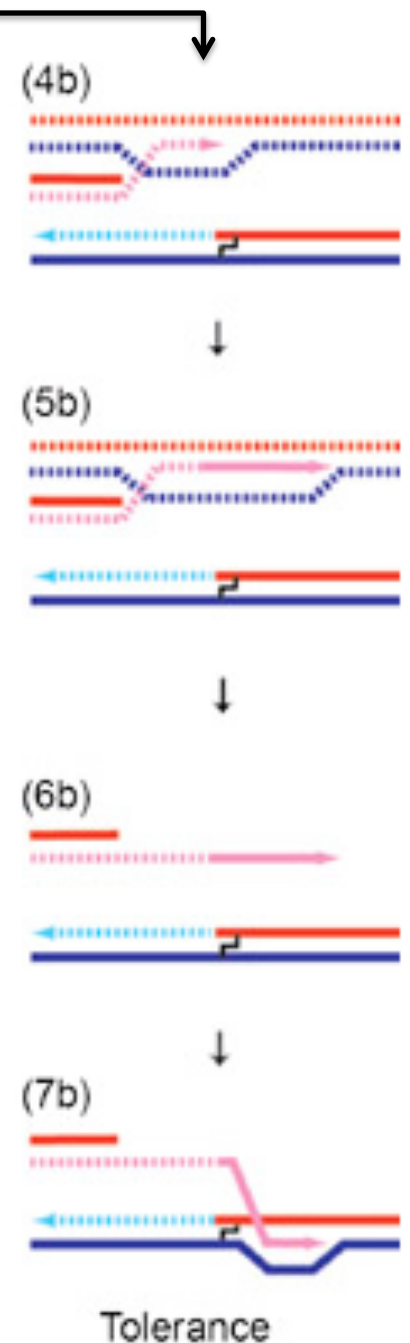
- <http://web.mit.edu/engelward-lab/animations/forkHR.html>

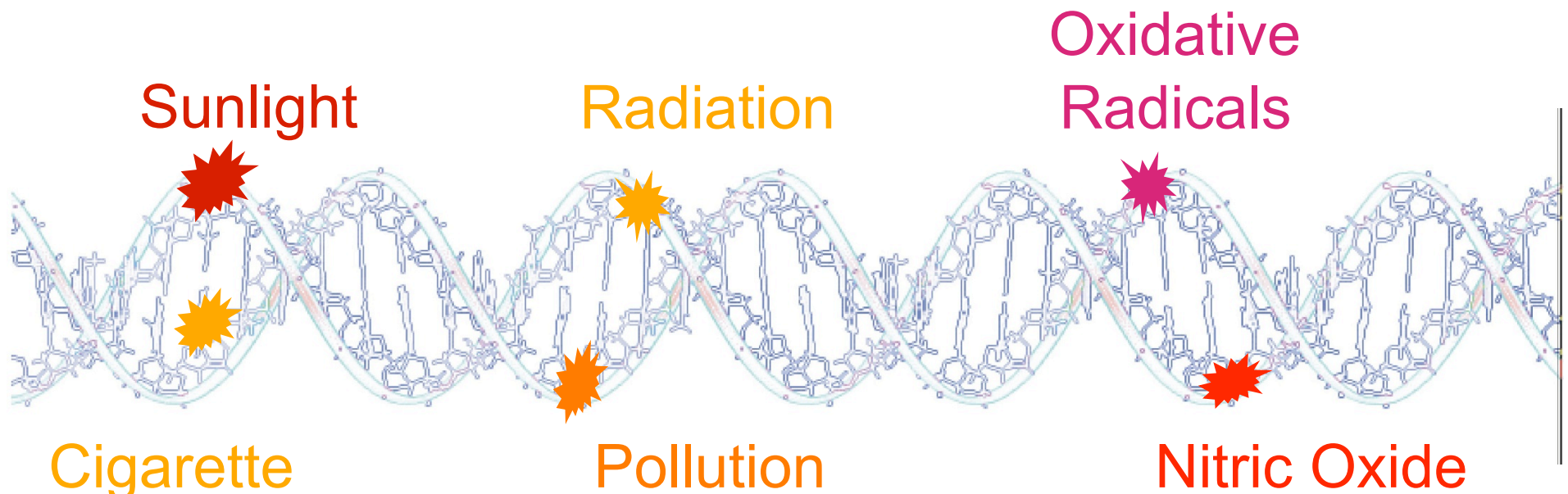


**Possible pathways to resolve replication forks stalled at interstrand crosslinks.** The stalled replication fork is recognized and cleaved in the leading-strand template to generate a one-sided DSB (steps 1-3).

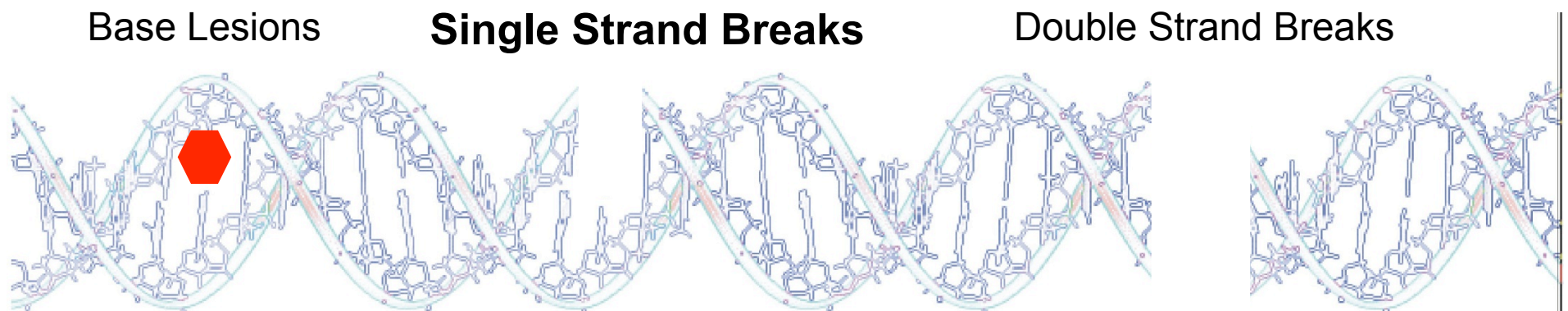
Introduction of a second incision on the other side of the ICL (step 4a) allows the lesion to flip out and to be bypassed by TLS (Trans lesion synthesis) (green line). The DSB is processed to form a 3'-OH ending single-stranded tail (step 5a) and to initiate DNA strand invasion (step 6a). The replication fork is restored (steps 7a) and the lesion is bypassed by TLS (green line).

The DSB can also initiate DNA strand invasion using the homolog as a template (step 4b). DNA is synthesized across the lesion region (step 5b), disengaged (step 6b) and reinvasion of the sister chromatid behind the lesion site can lead to restoration of the replication fork and tolerance of the lesion (7b; the step from D-loop to recovered fork are not drawn and equivalent to Figure 2A, steps 3a-4a-5a).



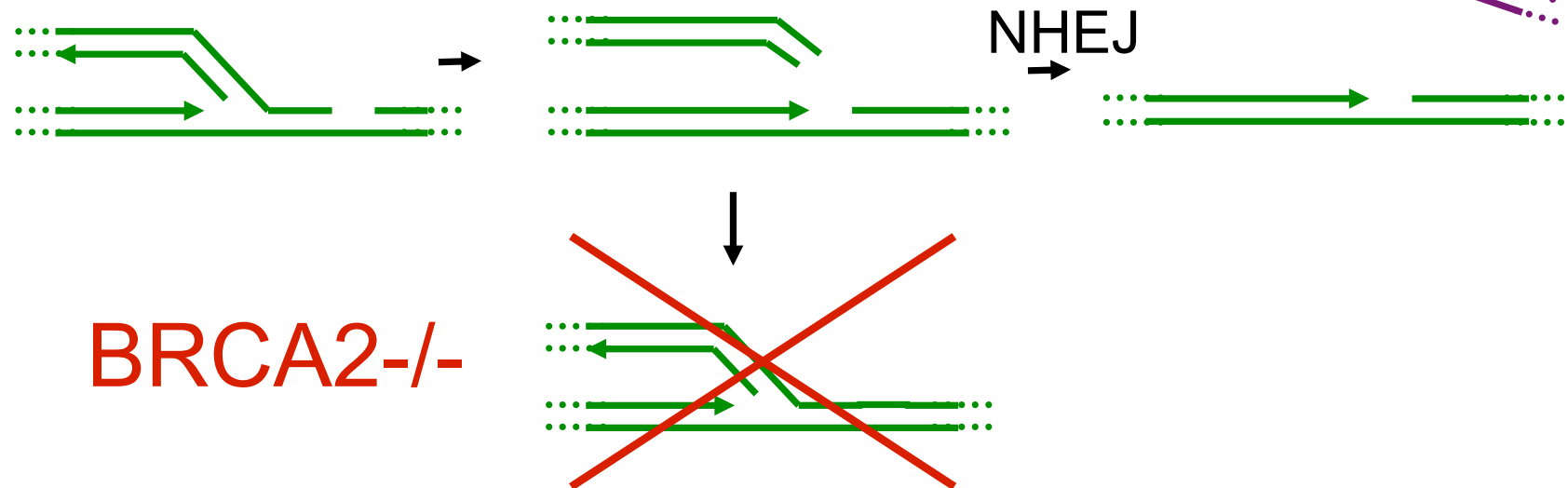


After damage, what might happen?



# BRCA2 is critical for repair of broken forks

What's bad about repairing fork breakdown with NHEJ?



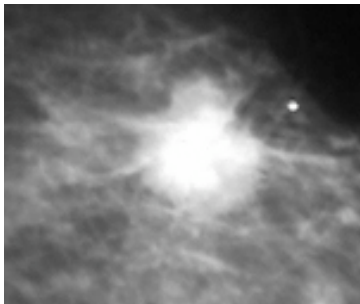


# Misrepair

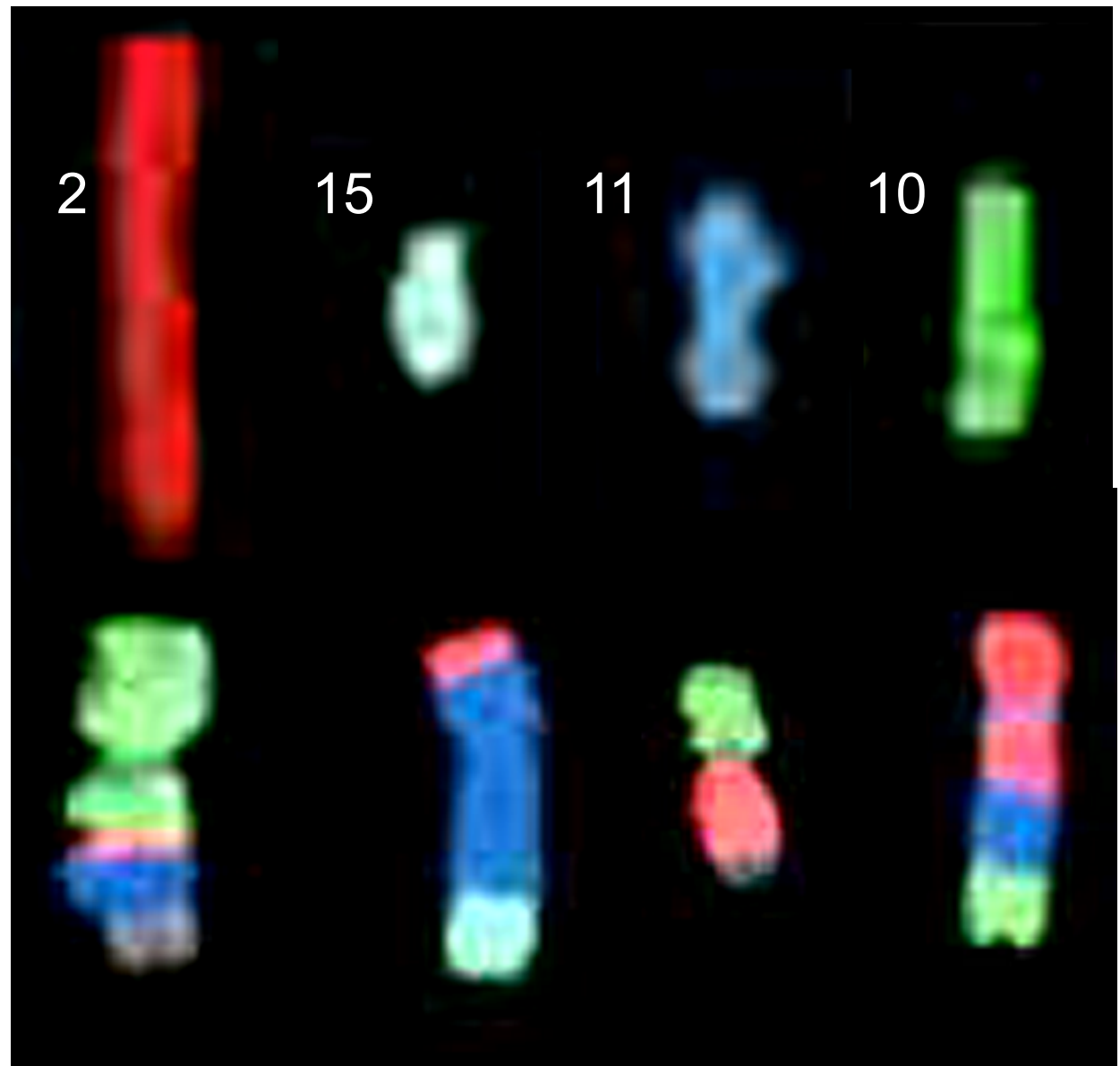
Normal Human  
Chromosomes



*BRCA2* -/-  
Chromosomes

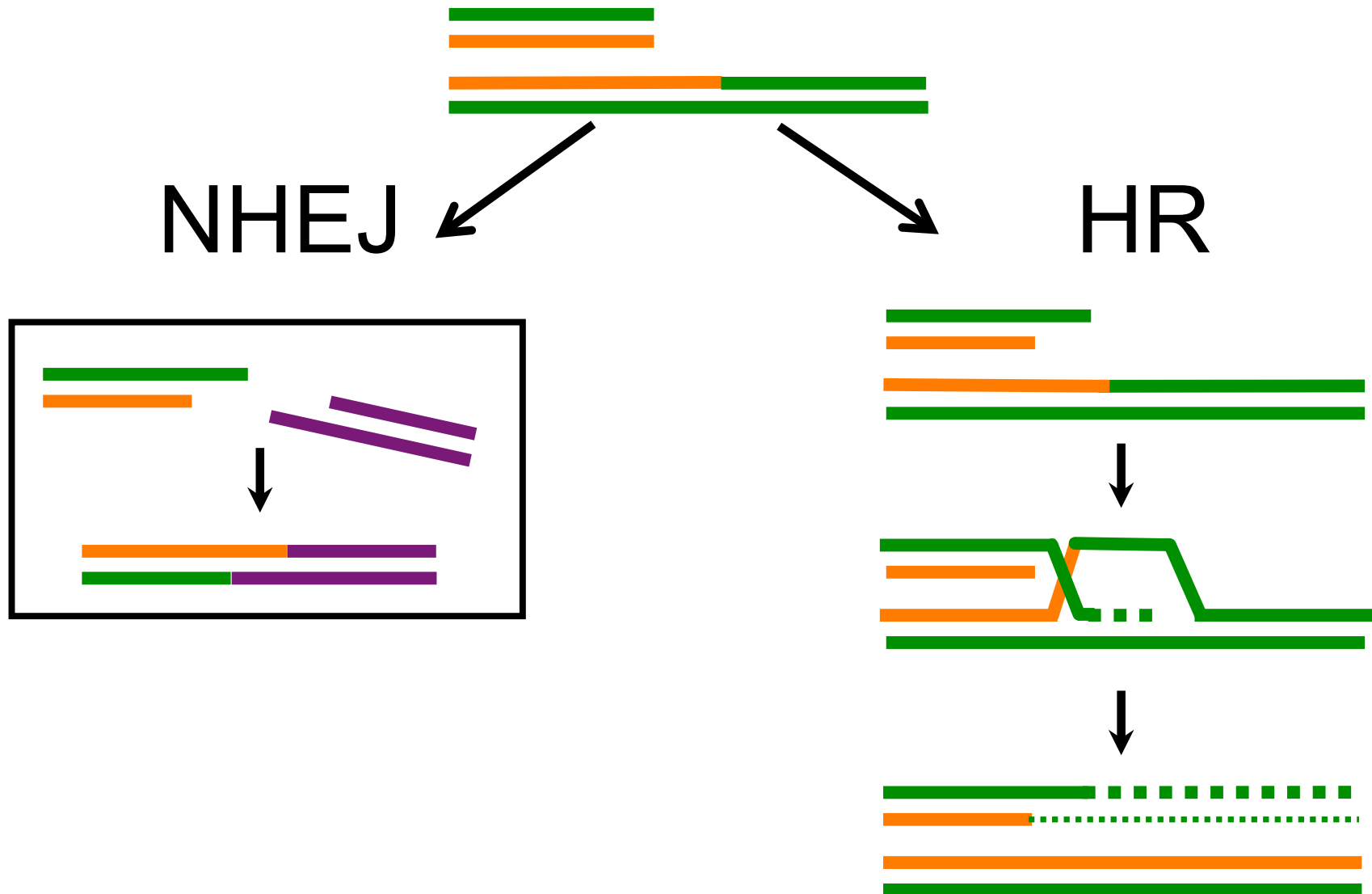


[www.rctradiology.com](http://www.rctradiology.com)



Raw data from Grigorova *et al.*, Cytogen. and Gen. Res. 104:333-340 (2004)

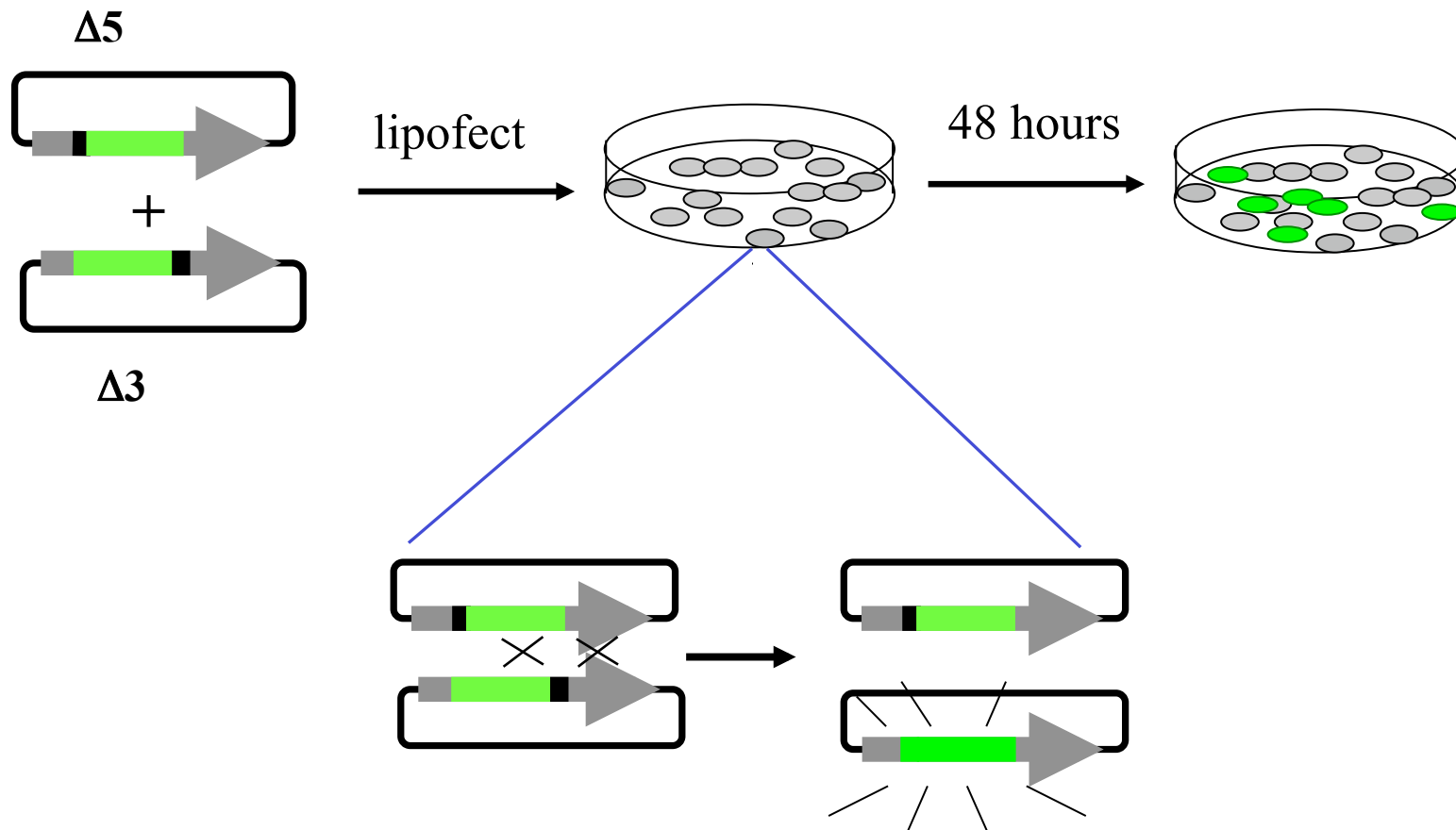
# Broken Fork Repair





# **Your Assay for Homologous Recombination**

# A Plasmid-Based Assay for Homologous Recombination in Mammalian Cells



- **About the experiments in Mod1**

- How does DNA damage cause mutations?
- How is recombination used to fix double strand breaks?
- Overview of the experiments you will be doing

- **Restriction Enzymes**

- Basics restriction enzymes

- **Anticipating Potential Problems & Pitfalls**

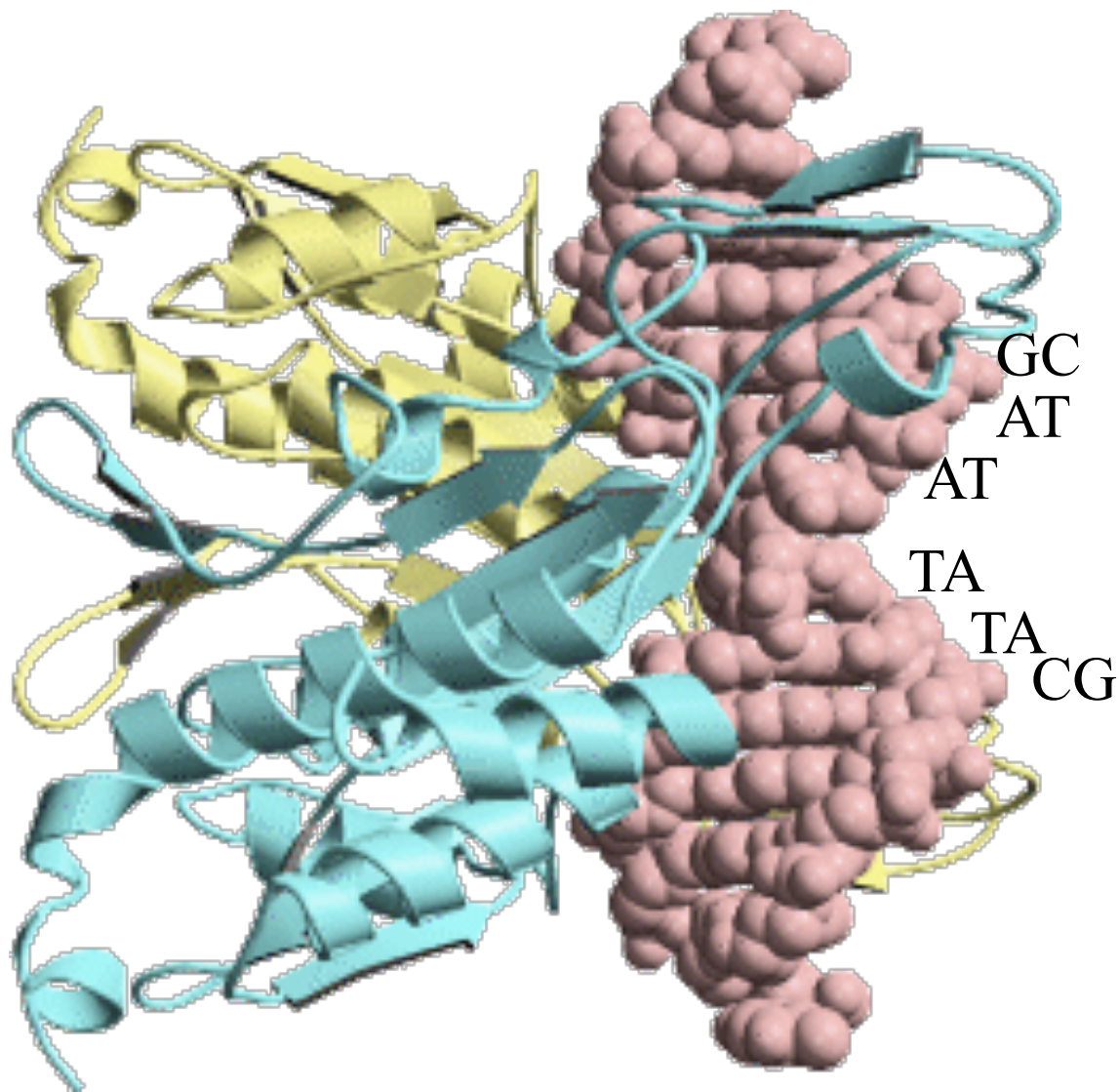
- What controls are needed and why?

# Overview of the Experiments in Mod1

Where you are,  
and where you are going

# Restriction Enzymes

- Where they come from?
  - Bacteria
- What do they do?
  - Chop up the DNA of infecting phages
- How do cells protect themselves?
- How can we use them?



**EcoRI**

Image from: Rosenberg, J. M. Curr. Opin. Struct. Biol. 1: 104-110 (1991)

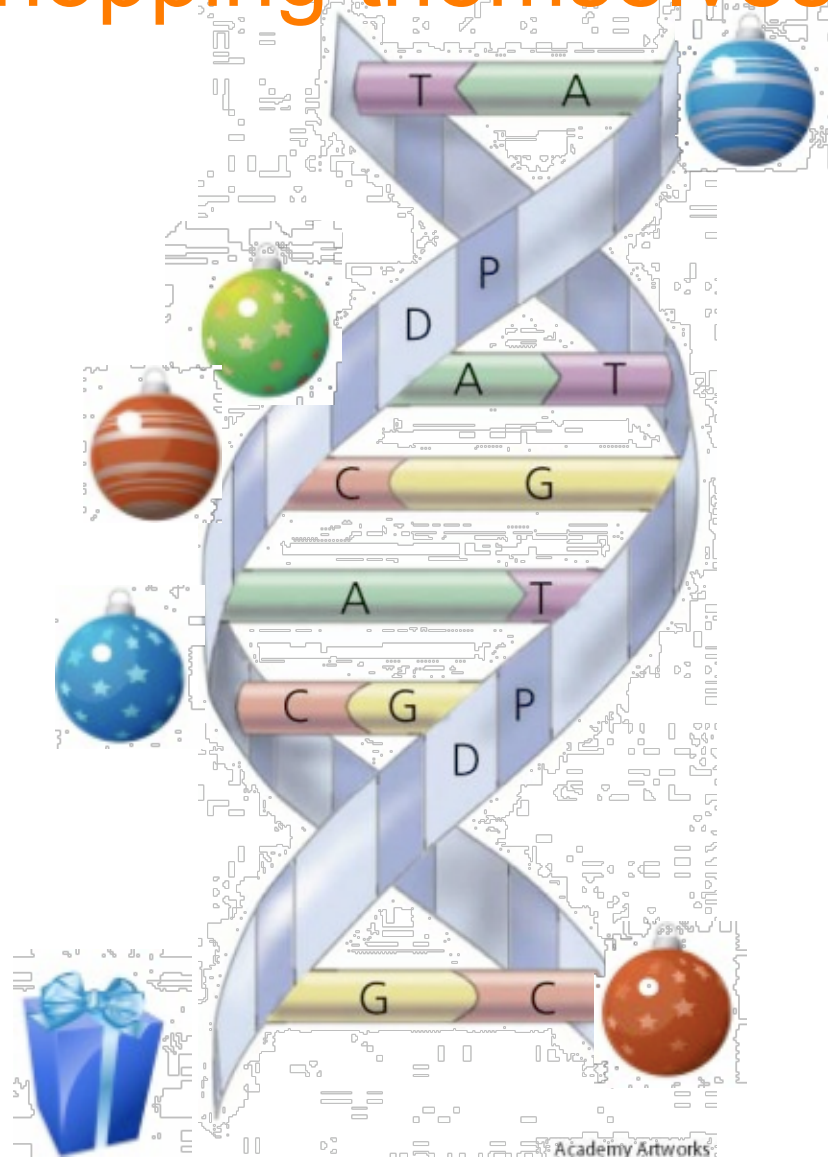
5' - G A A T T C - 3'  
3' - C T T A A G - 5'

**EcoRI**



5' - G                      A A T T C - 3'  
3' - C T T A A                      G - 5'

How do bugs keep from  
chopping themselves up?





# “Cognate Methyltransferases”

*M.HaeIII*

5' –GG**C**–3'

3' –C**C**GG–5'

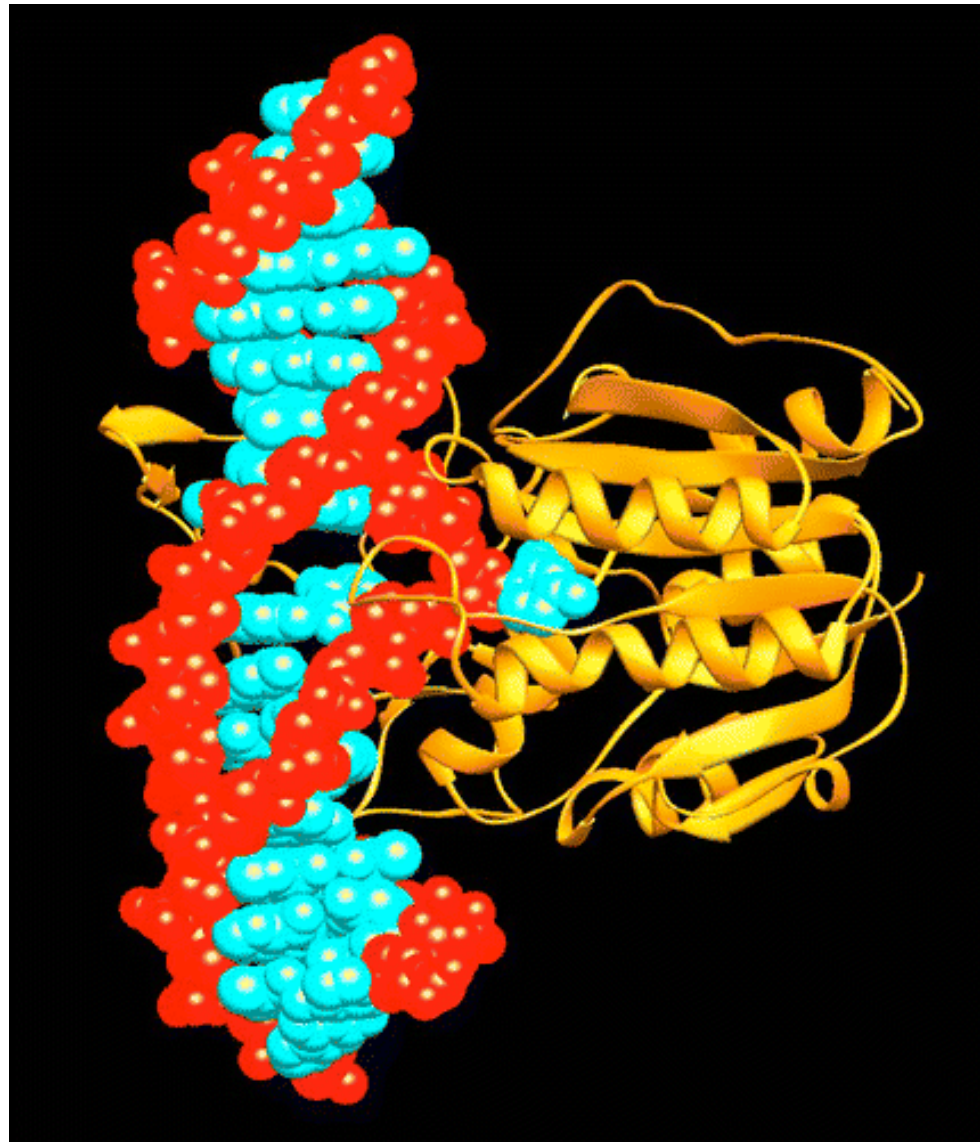
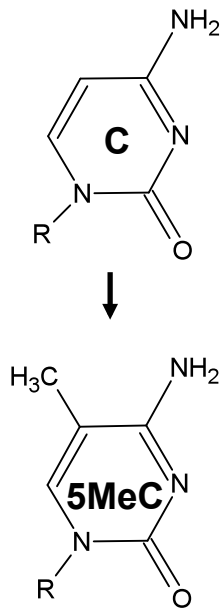
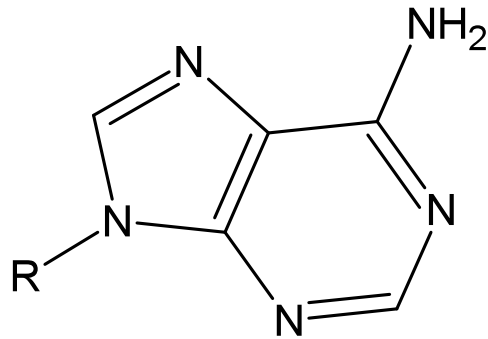


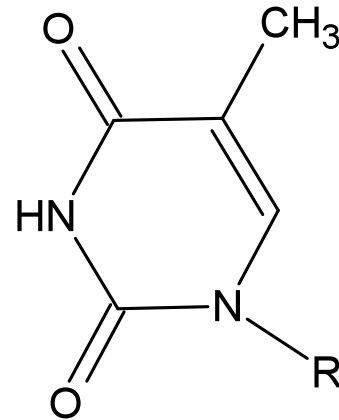
Figure from R. Roberts, Annual Review of Biochemistry (1998) 67: 181-198.

# Who's Who?

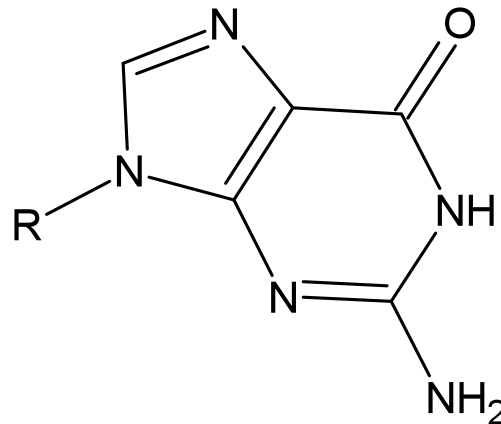
Adenine



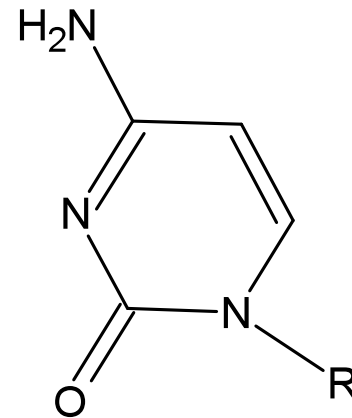
Guanine

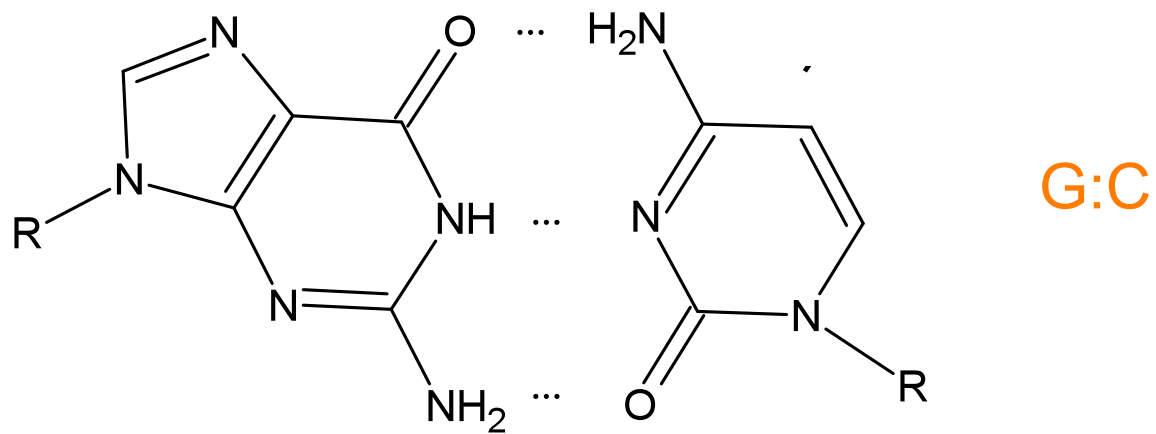
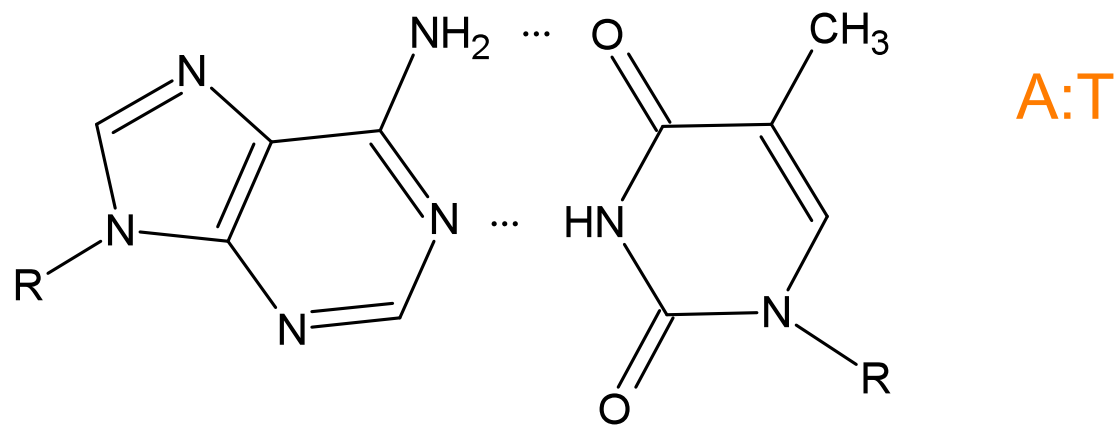


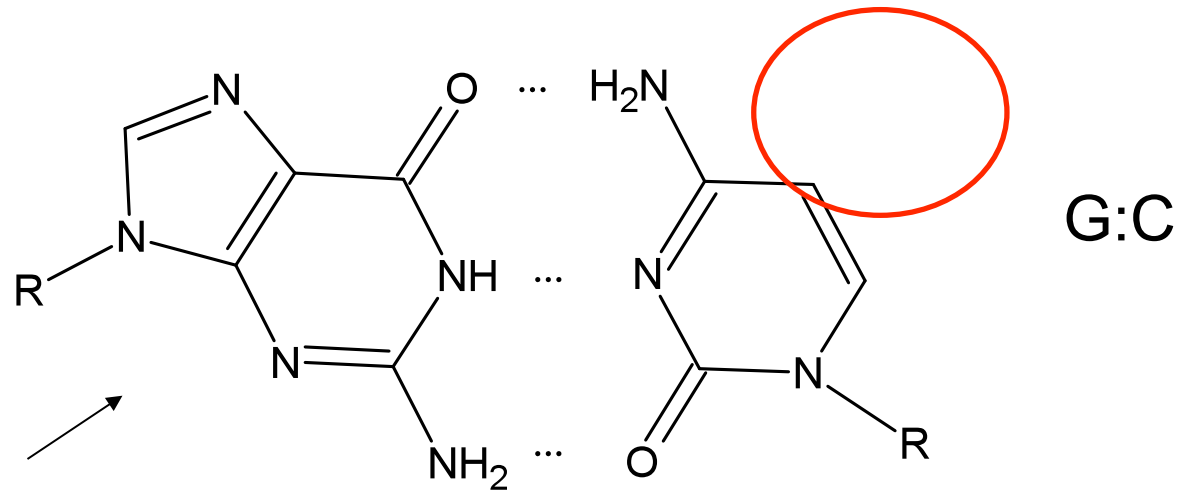
Thymine



Cytosine

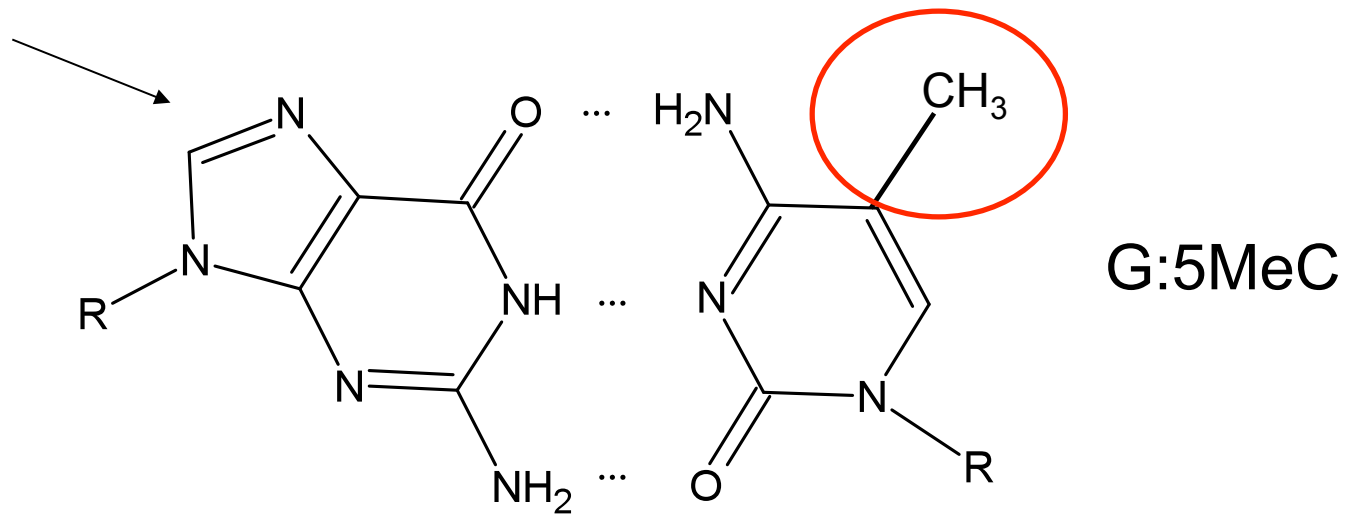






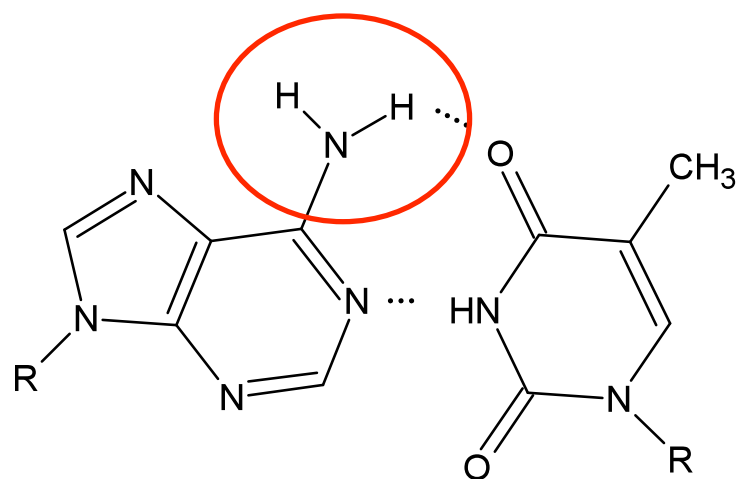
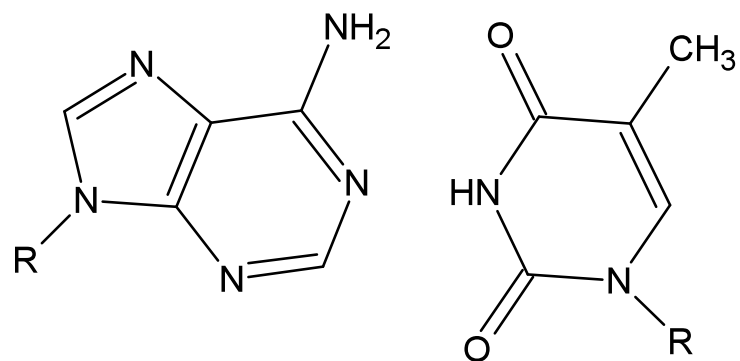
G:C

Proper  
G:C Base  
Pairs

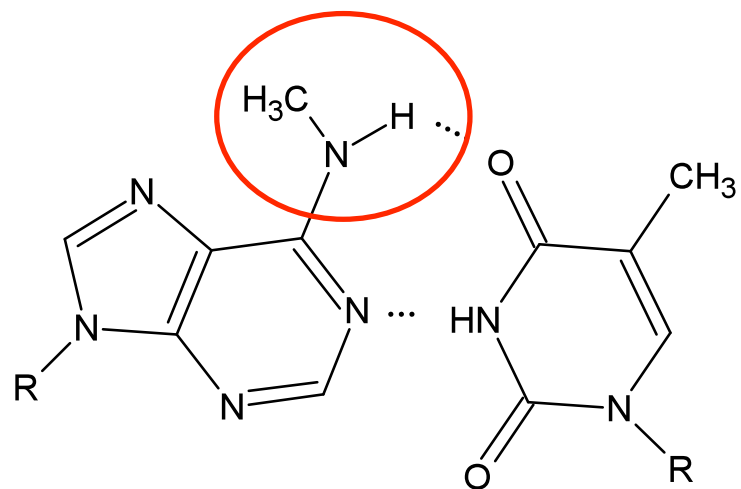


G:5MeC

Proper  
A:T Base  
Pairs



6MeA



# On a practical level...

## Using Restriction Enzymes

- Different kinds of restriction enzymes (blunt/distal)
- Shared recognition sequences
- Shared overhangs
  
- Buffer conditions
- Storing and diluting your restriction enzymes
- Specificity (potential pitfalls!)
- Lack of activity (host cells & potential pitfalls)

***Double check you're using the enzyme you think you are using!***

# Construction of the $\Delta 5$ Plasmid

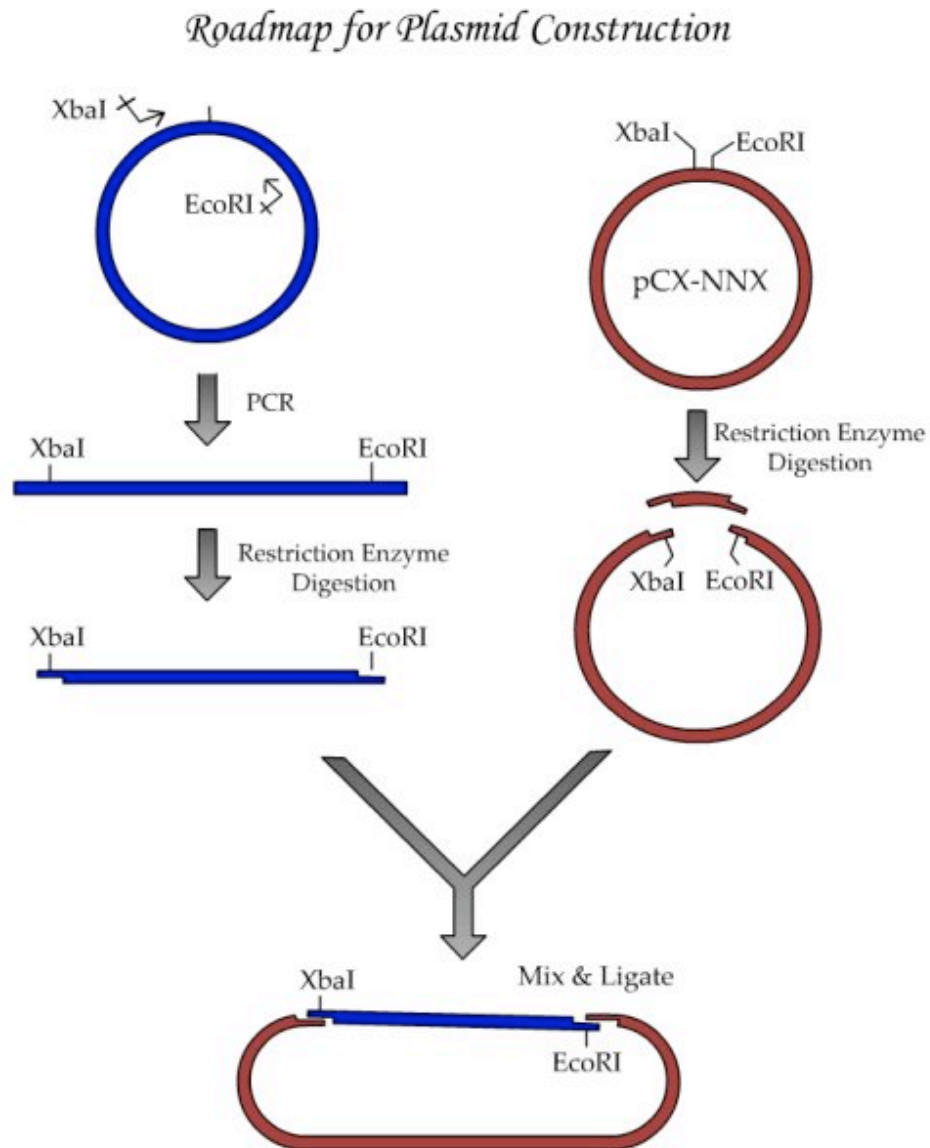


Figure by Justin Lo

# Anticipating Problems & Pitfalls:

What might go wrong in your experiment?

Incomplete Reactions

Controls: How can you tell if your DNA has actually been cut?



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- Basics restriction enzymes

- **Anticipating Potential Problems & Pitfalls**

- What controls are needed and why?